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Using mechanisms of hydrolysis and sorption to reduce siloxanes occurrence in biogas of anaerobic sludge digesters

Georgia Gatidou^a, Olga S. Arvaniti^a, Athanasios S. Stasinakis^{a*}, Nikolaos S. Thomaidis^b, Henrik R. Andersen^c

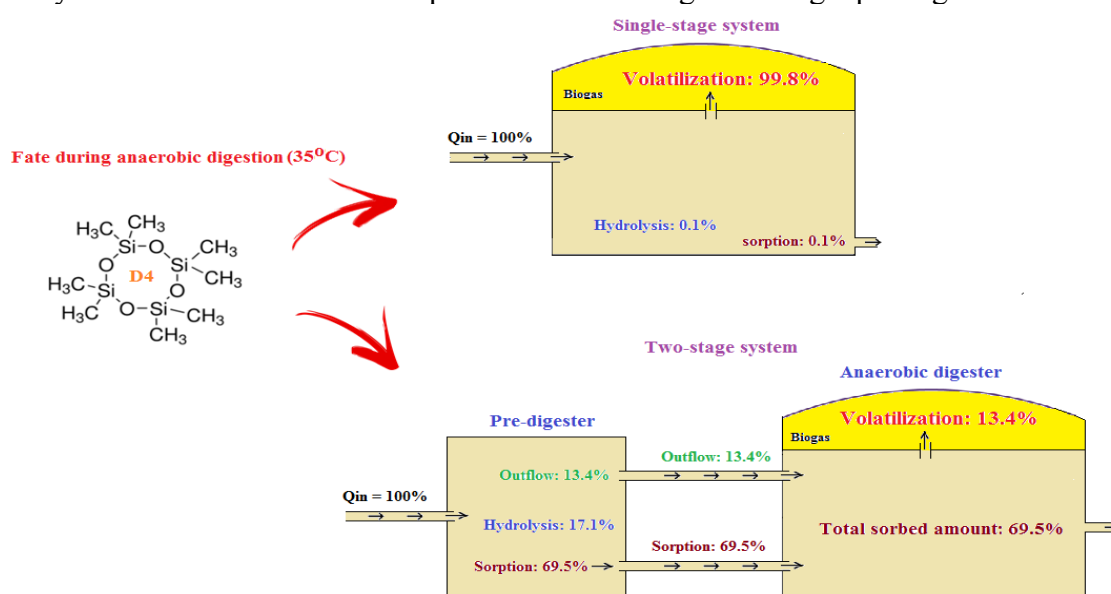
^aDepartment of Environment, University of the Aegean, 81100 Mytilene, Greece. *Corresponding author. Tel.: +30 22510 36257; fax: +30 22510 36206. E-mail address: astas@env.aegean.gr

^bDepartment of Chemistry, National and Kapodistrian University of Athens, 15771 Athens, Greece

^cDepartment of Environmental Engineering, Technical University of Denmark, Miljøvej 115, 2800 Kgs. Lyngby, Denmark

Highlights

- Hydrolysis and sorption constants for sludge (K_d) determined for cyclic siloxanes
- Hydrolysis increased with temperature and decreased with size of cyclic siloxanes
- Sorption increased with molecule size with $\text{Log}K_d$ values from 1.46 to 3.84
- During single-stage anaerobic digestion, > 91% of siloxanes end up in biogas
- Cyclic siloxanes volatilization predicted to be mitigated using a pre-digester



ABSTRACT Hydrolysis of hexamethylcyclotrisiloxane (D3), octamethylcyclotetrasiloxane (D4), decamethylcyclopentasiloxane (D5), dodecamethylcyclohexasiloxane (D6) and dodecamethylcyclohexasilane (D6_silane) and their sorption to digested sludge was studied in batch experiments. Hydrolysis was affected by the type of the compound and the applied temperature, while the relevant half-life values ranged between 0.07 ± 0.01 d (D3, 55 °C) and 48.4 ± 17.1 d (D6_silane, 4 °C). D5 showed the greatest affinity for sorption to digested sludge ($\text{log}K_d$: 3.84 ± 3.42), the lowest $\text{Log}K_d$ value was found for D3 (1.46 ± 0.95). Prediction of investigated compounds' fate in a single-stage anaerobic digestion system indicated that volatilization seems to be the major fate in both mesophilic and thermophilic conditions. The addition of a pre-digester with 3 d retention time would significantly decrease the expected concentrations of all siloxanes in biogas, enhancing their removal through hydrolysis and sorption to sludge.

Keywords: micropollutants; hydrolysis; sorption; fate; sludge treatment

Introduction

Siloxanes consist of a backbone of silicon and oxygen atoms which are alternated with organic side-chains attached to each silicon atom. These compounds can be found in linear, cyclic or tetrahedral formation with organic groups (Allen et al., 1997). Their physicochemical properties are noteworthy since they are hydrophobic and volatile at the same time depending on their molecular weight. The longer the chain length the lower the water solubility. Furthermore, these compounds are characterized by high stability, biocompatibility, surface activity and lubricating properties (Allen et al., 1997). Siloxanes can undergo hydrolysis in the environment. According to Durham (2005, 2006) this process is relatively slow in freshwaters under environmentally realistic conditions. Half-lives of octamethylcyclotetrasiloxane (D4) and decamethylcyclopentasiloxane (D5) have been estimated to be equal to 16.7 d and 315 d in freshwater at pH 7 and 12 °C, while the rate of hydrolysis was dependent on the pH and temperature. Among different siloxanes, D4 is toxic to sensitive aquatic organisms presenting long-term NOEC lower than 10 µg L⁻¹. D5 does not exhibit adverse effects on fish and water flea at concentrations as high as 17 µg L⁻¹ (Wang et al., 2013a) but shows considerable persistence and bioaccumulation (Sparham et al., 2008). Cyclic siloxanes have been also incriminated as potential endocrine disruptors, while they have been implicated in connective tissue disorders, adverse immunologic responses, and liver and lung damage in animal studies (Bletsou et al., 2013).

Due to their rather unusual properties, siloxanes are widely used in many applications such as antifoaming agents, coatings, shampoos, cosmetics (de Arespachaga et al., 2015) and as a result they end up in sewage treatment plants (STPs). Occurrence of siloxanes at ppb and ppm levels in wastewater and sludge, respectively, has been reported worldwide (Wang et al., 2013b; Bletsou et al., 2013; Van Egmond et al., 2013; Liu et al., 2014) and their fate during activated sludge process has been described (Tan et al., 2017; Xu et al., 2013; Wang et al., 2015a; Wang et al., 2015b). Among the different processes that determine the removal of compounds in STPs, volatilization in the aeration tank and adsorption to sludge are the most important pathways for the elimination of siloxanes (Wang et al., 2015a).

Certain semi-volatile siloxanes have also been detected in biogas produced by anaerobically digested sludge in concentrations high enough to damage the gas motors used to convert biogas to electricity (Dewil et al., 2006; Rasi et al., 2010; Raich-Montiu et al., 2014). Despite the frequent detection of siloxanes in sludge and produced biogas, so far, their fate during sludge anaerobic digestion has not been investigated. Additionally, no information is available for the hydrolysis of these compounds at different temperatures used during anaerobic digestion (35, 55 °C), as well as for their sorption potential in anaerobically digested sludge.

Based on the above, the main objectives of the present study were to investigate hydrolysis of selected cyclic siloxanes (hexamethylcyclotrisiloxane, D3; D4; D5; dodecamethylcyclohexasiloxane, D6; dodecamethylcyclohexasilane, D6_silane) (Table 1) under different temperatures including those typically used in mesophilic and thermophilic digestion (4 °C, 20 °C, 35 °C and 55 °C), and to determine sorption distribution coefficient (K_d) of these compounds in anaerobically digested sludge. Determination of investigated compounds in dissolved and solid phase was accomplished by applying a modified and fully validated previously developed (Bletsou et al., 2013) liquid-liquid extraction - gas chromatography - mass spectrometry method (LLE - GC-MS). A model was developed to predict the fate of investigated compounds during single-stage mesophilic and thermophilic anaerobic sludge digestion and to investigate the contribution of hydrolysis, sorption and volatilization to their elimination. Finally, a two-stage system was proposed to reduce the concentrations of investigated siloxanes in biogas.

2. Materials and Methods

2.1. Analytical standards and reagents

High purity analytical standards of D3, D4, D5, D6, D6_silane and tetrakis (trimethylsiloxy)-silane (M4Q) (all >95% purity) were purchased from Sigma-Aldrich (Seelze, Germany). M4Q was used as surrogate standard for the analysis. All organic solvents including ethyl acetate, methanol, pentane, dichloromethane and isooctane were supplied by Merck (Frankfurt, Germany) and were of high purity (>95%). KH_2PO_4 , Na_2HPO_4 , CaCl_2 and $\text{MgSO}_4 \times 7\text{H}_2\text{O}$ were purchased by Fluka (Buchs, Switzerland). Distilled water was provided by a MilliQ system (Milli-Q Gradient A10, Molsheim, France).

Standard stock solutions of individual compounds were prepared both in pentane and methanol at 1000 mg L^{-1} and stored in the dark at -18°C . Mixtures of investigated analytes were prepared both in pentane and methanol at concentrations of 10 and 100 mg L^{-1} . These mixtures were used to regularly prepare working standard solutions for calibration and spiking experiments.

2.2. Hydrolysis experiments

Hydrolysis experiments were conducted to investigate the role of this mechanism on the examined siloxanes. Phosphate buffer solution was prepared in Milli-Q water according to OECD protocol 111 in order to adjust pH at 7.0 ± 0.1 (OECD, 2004). The buffer solution was also enriched with small amounts of CaCl_2 and $\text{MgSO}_4 \times 7\text{H}_2\text{O}$, 0.03 g L^{-1} and 0.02 g L^{-1} , respectively. A known volume of buffer solution (43 mL) was placed in amber glass vials and small amounts of investigated compounds ($43 \text{ }\mu\text{L}$ in methanol) were added in each flask to give a final concentration of $100 \text{ }\mu\text{g L}^{-1}$ for each siloxane, except D3 for which the final concentration was chosen to be $300 \text{ }\mu\text{g L}^{-1}$ due to its higher quantification limit. Spiking was performed under the water surface and headspace was avoided in the vials to avoid volatilization. Experiments were made at four different temperatures ($4.0 \pm 1.0^\circ\text{C}$, $20.0 \pm 1.0^\circ\text{C}$, $35.0 \pm 1.0^\circ\text{C}$ and $55.0 \pm 1.0^\circ\text{C}$). Flasks were wrapped in aluminium foil to prevent photodegradation. Samples were taken at selected time intervals (3, 6, 15 and 24 h for D3 and 3, 7, 12, 16, 20 and 24 d for all the other compounds). Control runs were also performed at the same experimental conditions and samples were taken at t equal to 0 and 3 h to estimate the impact of spiking procedure and sorption onto flask on siloxanes removal. All experiments were conducted in triplicates.

2.3. Sorption experiments

Sorption experiments were performed using freeze-dried mesophilic anaerobically digested sludge taken from the Athens STP (Greece) which was prepared according to previous study (Arvaniti et al., 2014). Briefly, the preparation of the sludge was as follows: washing three times using tap water, centrifugation to remove water soluble constituents and freezing at 18°C for 24 h. Thereafter, the sludge was gently freeze dried to preserve its structure, sterilized by heating at 103°C for minimum 3 h and stored at 4°C . The characteristics of digested sludge are presented in Table S1.

To determine K_d values of the investigated compounds, batch experiments were conducted for a range of initial concentrations of each compound (50, 75, 100, 125, 150 and $200 \text{ }\mu\text{g L}^{-1}$). 1 g L^{-1} sludge and phosphate buffer solution (43 mL) were added in each flask, in order to avoid headspace. Batch experiments were conducted at six different concentrations (150, 225, 300, 375, 450 and $600 \text{ }\mu\text{g L}^{-1}$). A stir bar was added in every glass flask, in order to achieve homogeneous mixing of the samples. Flasks were covered with aluminium foil to prevent photodegradation, agitated at 120 rpm on a shaking plate and samples were taken at the end

of the experiment (24 h) for the analysis of compounds in the dissolved phase. For D3 and D4, K_d values were determined by analysing both solid and dissolved phase due to their higher instability (OECD, 2006).

Control runs without sludge were performed at the same experimental conditions to estimate the impact of other processes (e.g., spiking procedure) on siloxanes removal. Furthermore, unspiked samples were analyzed to quantify background concentration of each analyte in sludge. All the experiments were performed in triplicates at 35.0 ± 1.0 °C and pH 7.0 ± 0.1 . Further details on the experimental design are described in Table S2.

2.4. Siloxanes analysis

2.4.1 Samples pretreatment

The analytical procedure for the extraction of wastewater and sludge samples was based on previously developed method (Bletsou et al., 2013), which was applied after several modifications and full validation.

For wastewater, 10 mL of filtered sample was transferred into a 50 mL Eppendorf tube and a known amount of M4Q (300 ng prepared in methanol) was added before extraction. After 20 min of equilibration, investigated analytes were extracted for 1 min by LLE with 10 mL of pentane. This solvent was chosen among others which were tested (pentane, hexane, heptane, octane) since resulted in the less contribution to the total background. Samples were extracted two more times with 2.5 mL mixture of pentane:dichloromethane (50:50, % v/v) and 2.5 mL of mixture pentane:ethyl acetate (50:50, % v/v). All three supernatants were transferred into a 15 mL Eppendorf tube. The combined organic layers were evaporated to a final volume of 500 μ L under a gentle stream of nitrogen gas (N_2). Finally, the eluents were transferred into a vial containing 500 μ L of pentane and subjected to GC-MS analysis.

Regarding sewage sludge, 85 mg of dewatered sludge was transferred into a 15 mL Eppendorf tube. Samples were spiked prior to extraction with 300 ng of M4Q (prepared in methanol) and allowed to equilibrate for 20 min at -18 °C. Then, 5 mL of pentane and 1 mL of isooctane were added into tube and shaken for 22 hours at 300 rpm in the dark. The supernatant was collected into a 15 mL Eppendorf tube after centrifugation at 5000 rpm for 10 min. Samples were extracted three more times using 5 mL of pentane, 2.5 mL of pentane:dichloromethane (50:50, % v/v) and 2.5 mL of pentane:ethyl acetate (50:50, % v/v). Every extraction step was lasted 1 h and followed by centrifugation at 5000 rpm for 10 min. Finally all supernatants were combined and followed the same procedure as aforementioned for the dissolved phase samples of wastewater.

2.4.2 GC-MS Analysis

Analysis was performed on an Agilent 7890A GC interfaced with an Agilent 5975C MS slightly modifying the method of Bletsou et al. (2013). Separation was achieved by a DB5MS capillary column (60 m) with a film thickness of 0.25 μ m and internal diameter of 0.32 mm (Agilent Technologies, USA). Two μ L of extract were injected using a 768313 Agilent autosampler (Agilent Technologies, USA) in a splitless mode at 200 °C and helium was used as a carrier gas at a flow rate of 1.0 mL min⁻¹. The oven temperature was programmed from 40 °C (kept for 1 min); it was then increased to 220 °C at a rate of 20 °C min⁻¹ and from 220 to 280 °C at a rate of 25 °C min⁻¹, with a post-run time of 5 min at 300 °C. The MS was operated in an electron-impact ionization mode (EI) at 70 eV, and the data were acquired by selected ion monitoring (SIM). To evaluate the best mass spectral data for each siloxane, injection of every individual compound was performed in full scan (mass range from 50-500) and the most abundant ions were selected from its spectrum. The chosen ions for SIM mode were 207, 191, 96 for D3, 281, 265, 349, 133 for D4, 355, 267 for D5, 348, 201, 215 for D6,

281, 147, 369 for D6-silane and 341, 429, 325 for M4Q. The selected ions are in agreement with those reported in the open literature (Bletsou et al., 2013). A typical chromatogram is presented in Figure S1.

The main difficulty of siloxanes analysis is background contamination. Based on previous report (Tran et al., 2015), a number of actions were taken in order to control and minimize sources of contamination. To minimize this effect, all glassware was placed in an oven at 100 °C until use. As the GC vial caps were not free of silicon, vials were capped with aluminium foil which previously was baked at 100 °C for 24 h. Furthermore, the analysts refrained from using hand lotions and other personal care products that contained siloxanes. The procedural blank was analyzed with every 10 samples and background levels of siloxanes were taken into account during interpretation of the results. Prior to instrumental analysis, pentane (reagent blank) was injected into GC-MS, until the background levels of siloxanes became stable. In addition, standard solution and spiked sample were injected after every 10 sample injection to evaluate the drift in instrumental sensitivity. Pentane was injected after every 10 samples as a check for background contamination and carry-over.

2.5 Validation of the analytical methods

Validation of the analytical methods included analytical methods calibration, determination of limits of detection (LODs), assessment of precision and evaluation of trueness for both dissolved and particulate phase samples.

Calibration of the GC-MS system was performed using eight different concentrations ($n=3$) in the range of 10-1000 $\mu\text{g L}^{-1}$. Matrix matched calibrations were also performed using fortified samples in the range of 10-1000 ng for both types of samples. For the determination of the analytical LOD, 10 mL of wastewater were spiked with 7 ng of the investigated siloxanes, whereas 85 mg of dry sludge spiked with 10 ng. LOD was calculated as three times the standard deviation of nine independent replicate analyses. Intra-day precision data was obtained by analyzing six samples ($n = 6$) spiked at a level of 30 $\mu\text{g L}^{-1}$ of each compound. Inter-day precision data was obtained by analyzing six samples ($n = 6$), spiked at the same level, at three different days ($k = 3$). During these experiments sludge samples were spiked with 300 ng of siloxanes. Recovery studies were performed at three fortification levels both in wastewater (10, 30 and 80 $\mu\text{g L}^{-1}$) and sludge (100, 300 and 800 ng), in order to evaluate the trueness of the method.

2.6 Calculations

For each siloxane, the degree of removal efficiency in hydrolysis experiments was calculated using Eq. (1):

$$\text{Removal}(\%) = \frac{C_0 - C_w}{C_0} \times 100 \quad (1)$$

Where, C_0 is the measured concentration of each siloxane ($\mu\text{g L}^{-1}$) at $t = 0$ in flasks (control run) and C_w is the analyte concentration ($\mu\text{g L}^{-1}$) in the dissolved phase at different time intervals.

Moreover, the obtained data from hydrolysis batch experiments were described by the pseudo first-order kinetics, (Eq. 2):

$$C_t = C_0 e^{-k_{\text{hydrolysis}} t} \quad (2)$$

Where C_t and C_0 are the dissolved investigated compound concentrations in batch experiment at time t and $t = 0$, respectively, ($\mu\text{g L}^{-1}$), $k_{hydrolysis}$ is the hydrolysis coefficient (h^{-1} or d^{-1}) and t is the time at the end of experiments (hours or days).

Sludge-water distribution coefficients (L Kg^{-1}) of investigated compounds were estimated from sorption experiments according to Eq. 3:

$$K_d = \frac{C_s}{C_w} \quad (3)$$

Where, C_s and C_w are the equilibrium concentration of each investigated compound in sludge (ng Kg^{-1}) and water (ng L^{-1}), respectively. As mentioned in Paragraph 2.3, C_s concentration was measured for D3 and D4 to solid phase samples, whereas for the other investigated compounds C_s concentration was calculated using Eq. (4),

$$C_s = \frac{C_0 - C_w}{SS} \quad (4)$$

Where, C_0 is the measured concentration of each analyte in the water phase at $t = 0$ in flasks with no sludge addition (ng L^{-1}) and SS is the concentration of sludge suspension used (Kg L^{-1}).

Having in mind that investigated compounds are considered resistant to biodegradation (Graiver et al., 2003) and assuming that sorbed compounds are not subjected to hydrolysis or volatilization, Equations 5 and 6 were used to predict their fate in a single-stage mesophilic or thermophilic anaerobic digester:

$$M_{in} = M_{hydrolysis} + M_{sorption} + M_{volatilization} + M_{out} \quad (5)$$

Where M_{in} and M_{out} are the masses of investigated compounds in influents and effluents, respectively ($\mu\text{g d}^{-1}$), $M_{hydrolysis}$, $M_{sorption}$ and $M_{volatilization}$ are the masses of investigated compounds that are hydrolysed, sorbed and volatilised, respectively ($\mu\text{g d}^{-1}$).

$$C_{in}Q_{in} = (k_{hydrolysis}C_{out}V) + \frac{(X_T V K_d C_{out})}{SRT} + (Q_g H C_{out}) + (Q_{out}C_{out}) \quad (6)$$

Where C_{in} and C_{out} are the concentrations of investigated compounds in influents and effluents, respectively ($\mu\text{g m}^{-3}$), Q_{in} and Q_{out} are the flow rates in influents and effluents, respectively ($\text{m}^3 \text{d}^{-1}$), Q_g is the biogas flowrate ($\text{m}^3 \text{d}^{-1}$), $k_{hydrolysis}$ and K_d are the experimentally calculated hydrolysis and sorption constants, respectively, X_T is the concentration of suspended solids (g TSS L^{-1}), V is the volume of anaerobic digester (m^3), SRT is the sludge residence time in anaerobic digester (d) and H is the Henry's Law coefficient at 35°C or 55°C ($\text{m}^3 \text{liquid}/\text{m}^3 \text{gas}$).

It should be mentioned that siloxanes are neither classified as base or acid compounds, the biogas amount was calculated from the volatile suspended solids (VSS) reduction at a rate of 1 m^3 biogas per Kg VSS removed (Tchobanoglous et al., 2002), while Henry constants were estimated for 35°C and 55°C using Equation 7:

$$H = H_{25} \theta^{(T-25)} \quad (7)$$

Where H_{25} is the Henry's Law coefficient at 25 °C ($\text{m}^3 \text{ liquid m}^{-3} \text{ gas}^{-1}$), T is temperature at 35 °C or 55 °C and θ is the temperature correction factor (dimensionless). Value of θ was considered equal to 1.044, based on Gossett (1987) and TOXCHEM (2012). H_{25} was calculated using the Equation 8 (Kochetkov et al., 2001):

$$H_{25} = \frac{P_{25}}{RTS_{25}} \quad (8)$$

Where P_{25} is the vapour pressure at 25 °C (Pa), S_{25} is the water solubility at 25 °C (mole/m^3), R is the gas constant ($\text{Pa m}^3 \text{ K}^{-1} \text{ mole}^{-1}$) and T is the temperature (K).

The software GraphPad Prism 5 for Windows was used for data evaluation.

3. Results and Discussion

3.1 Validation of the analytical method

Satisfactory linearity was observed all along the tested area of concentration for all the compounds both during instrumental and matrix matched calibrations with coefficients of correlation (R^2) > 0.99. The obtained LODs varied from 2.2 (D4) to 4.6 (D6) ng mL^{-1} and from 0.1 (D4) to 1.8 (D5) ng mg^{-1} for wastewater and sludge, respectively (Table S3). These values are higher compared to other authors (Bletsou et al., 2013). However, it should be taken into account the lower preconcentration factors in the present study (only 10 mL of wastewater and 85 mg of sludge were extracted) compared to others. The precision experiments shown acceptable repeatability and reproducibility of the analytical procedures, both during the same day and between days (RSDs < 10.2%) (Table S4). Furthermore, satisfactory recoveries were observed for most of the tested siloxanes for both type of samples, ranging from 67% (D4) to 110% (D6_silane) with RSDs \leq 13% (Tables S5 and S6). For wastewater samples, D3 was the only compound for which low recoveries were observed (Table S5), while for sludge samples D5 shown very high recoveries (up to 158%) but with good repeatability (RSDs \leq 4.4%) (Table S6). In general, the results are comparable with those reported by other authors, although much lower amounts of wastewater and sludge sample were extracted (Bletsou et al., 2013; Sanchís et al., 2013). The low recovery of D3 has also reported by other authors (Sanchís et al., 2013) and it is due to the high vapour pressure of this compound.

3.2. Hydrolysis experiments

According to the results all tested siloxanes undergo hydrolysis. D3 was found to be the most sensitive among the five compounds (Figure 1a). About 70% of its initial concentration disappeared after 15 h even at 4 °C, while the elimination increased with the increase of temperature (90% loss at 55 °C in 15 h) since the rate of hydrolysis is temperature dependent with shorter half-lives at higher temperatures (Wang et al., 2013b).

Hydrolysis of other investigated compounds was much slower and it decreased with the increase in chain length (Figure 1b). D4 loss due to hydrolysis reached 70%, 95%, 96% and 97% at 4, 20, 35 and 55 °C, respectively, after 24 d of experiment. D5 reduction was evident after the 3rd day of the experiment and at the end of the experiment varied between 46% and 84% at 4 °C and 55 °C, respectively. As for D6, 48% of the compound was hydrolysed throughout the experiment at 4 °C, while its removal ranged up to 71% at 55 °C (24 d). Finally, D6_silane concentration loss varied from 32% (4 °C) to 56% (55 °C) during the experiment (24 d). Other authors have also reported lower hydrolysis rates regarding investigated compounds as their molecule increases (Environmental Agency 2009a, b, c). A

possible explanation for this “protection” against hydrolysis could be the effect of steric hindrance of the compounds.

First order kinetic equation was fitted to concentration profiles at each of the examined temperatures. The significance of the effect of temperature on the hydrolysis rate constant was analyzed by F-test in GraphPad Prism 5 with the null hypothesis being that the rate constant for degradation is similar at all the tested temperatures, for each siloxane, which revealed a statistical clear effect with $P = 0.0032$ for D6 and $P < 0.0001$ for the other compounds. The degradation rate constants were also significantly different between the investigated siloxanes at the four investigated temperatures with $P < 0.0001$ except for 4 °C there $P = 0.0248$.

The hydrolysis rate constant and the half-life of each compound were calculated with 95% confidence intervals (Table 2). The lowest half-life values were observed for D3 ranging between 0.07 d (± 0.01) at 55 °C and 0.27 d (± 0.07) at 4 °C, whereas D6_silane was found to be most stable with half-lives up to 48 d (± 17.1) at 4 °C.

Dow Corning (2005) reported a half-life of 1.04 d for D4 at pH 7 at 35°C which is comparable with the present value of 1.6 d. As for the other compounds, there is no hydrolysis data in the literature for similar experimental conditions. However, at pH 7 and 25 °C hydrolysis half-lives of 23 minutes and 71 days have been reported for D3 (Dow Corning, 2004) and D5 (Environment Agency, 2009b), respectively.

3.3. Sorption experiments

The sorption of the investigated compounds to anaerobically digested sludge was studied in flasks containing different concentrations of the target compounds at 35 °C. Sorption equilibrium constants were calculated for each siloxane using Equations 3 and 4. The results clearly distributed evenly in a linear plot of measured C_w against C_s and therefore no isotherm fittings were attempted. K_d values were calculated as the average of all the K_d determined ($n = 18$). As shown in Table 3, $\log K_d$ values varied from 1.46 (± 0.95) for D3 to 3.84 (± 3.42) for D5. This is the first study that report cyclic siloxanes sorption constants onto anaerobically digested sludge.

The sorption results are generally comparable with values published for other types of sludge. Bletsou et al. (2013) estimated $\log K_d$ values for D5 and D6 in primary and secondary sludge using full-scale monitoring data of a STP and reported values ranging between 1.83 (D3) and 4.42 (D5). Wang et al. (2015) determined $\log K_d$ values in primary and secondary sludge as high as 3.53 (D5) and 5.57 (D5), respectively. Xu et al. (2013) found slightly lower $\log K_d$ values in aerobic activated sludge which varied from 1.94 (D4) to 2.46 (D5). The variances observed among different studies are due to the different chemical characteristics of each type of sludge used (pH, organic carbon, cations). In the literature it has already been mentioned by several authors that the type of sludge affect the degree of micropollutants sorption and thus the values of K_d (Ternes et al., 2004; Hörsing et al., 2011; Arvaniti et al., 2012).

3.4. Prediction of siloxanes fate during single-stage anaerobic sludge digestion

The determined hydrolysis and sorption constants as well as values of the vapour pressure values taken from the literature (Table 1) were used to predict the contribution of hydrolysis, sorption and volatilization, respectively, for D3, D4, D5 and D6 during mesophilic and thermophilic anaerobic digestion. For this reason, Equation 6 was applied for single-stage anaerobic digesters operating at SRT of 17d and temperatures of 35 °C and 55 °C. For the operational parameters of the anaerobic digesters (volume, SRT, flowrates, VSS removal, biogas production), the anaerobic digester of Athens STP was used as model. All values of

different parameters used in Equation 6 as well as the operational characteristics of the anaerobic digesters are reported in Table S7. Equation 6 was not applied for D6_silane due to the lack of literature data for the vapour pressure value of this compound.

The results of the model are presented in Figure 2 as well as in Tables S8 and S9 (Scenario A). The mass balance shows that the major mechanism affecting the fate of investigated compounds for both mesophilic and thermophilic conditions is volatilization. More than 91% of all siloxanes seem to be transferred to biogas under mesophilic conditions, while this mechanism is further enhanced under thermophilic conditions, reaching 99.8% for D4. Despite the high hydrolysis constants of D3, its contribution to the removal of the investigated compound in a single-stage system was not of high importance, ranging up to 8.2% (35 °C). The contribution of hydrolysis for the other siloxanes did not exceed 0.2%. Small fractions of D5 (3.9%) and D6 (2.2%) seem to be sorbed onto sludge during mesophilic digestion, while even smaller sorbed amounts are estimated for thermophilic conditions. The role of this mechanism seems to be negligible for D3 and D4.

So far, the fate of the investigated cyclic siloxanes during anaerobic digestion has not been studied and the contribution of different mechanisms has not been investigated. Several previous articles suggested that D3, D4 and D5 were among the most prevalent siloxanes in biogas produced in STPs and biogas plants. Concentrations ranging between 40 and 420 $\mu\text{g m}^{-3}$ for D3, 30 to 10120 $\mu\text{g m}^{-3}$ for D4 and 100 to 15500 $\mu\text{g m}^{-3}$ for D5 have been reported (Rasi et al., 2010; Tansel and Surita, 2014; Raich-Montiu et al., 2014). Back-calculation of the results of this study shows that for the model anaerobic digester used and mesophilic conditions, concentrations equal to 670 $\mu\text{g m}^{-3}$, 728 $\mu\text{g m}^{-3}$ and 700 $\mu\text{g m}^{-3}$ are estimated for D3, D4 and D5 respectively, in biogas. These concentrations are the same order of magnitude with those reported in the literature.

To investigate the sensitivity of the mass balance based model to possible differences of the experimentally calculated constants of this study, two different scenarios (B, C) were tested and the results are given in Tables S8 (mesophilic digester) and S9 (thermophilic digester). Specifically, an increase of $k_{\text{hydrolysis}}$ values by 50% (Scenario B) would increase the contribution of this mechanism on D3 removal by 3.5% (mesophilic conditions) or 2.4% (thermophilic conditions), whereas no important effect would be noticed for the other compounds. On the other hand, an increase of K_d values by 50% (Scenario C) would not actually affect the behaviour of D3 and D4, while it would increase by 1.9% and 1.0% the contribution of sorption on D5 and D6 removal under mesophilic conditions, respectively (Table S8). A slight increase of sorbed amounts of these two compounds is also predicted for thermophilic conditions (Table S9). The aforementioned calculations indicate that a possible underestimation by 50% of hydrolysis and sorption constants in this study would not affect significantly the results regarding the fate of the investigated siloxanes during sludge anaerobic digestion. Future monitoring studies in full-scale anaerobic digesters should be performed to confirm the results of the prediction model developed in this article.

3.5. Use of a two-stage system for decreasing siloxanes concentration in biogas

The occurrence of siloxanes in biogas is responsible for commonly observed damages in the gas motors (Rasi et al., 2010; Raich-Montiu et al., 2014), resulting in increased cost for renovation or treatment of the produced biogas by activated carbon sorption (Soreanu et al., 2011). On the other hand, the short hydrolysis half-lives of D3 and D4 (<2d, Table 2) indicate that the operation of a pre-digester tank with low hydraulic residence time (HRT) could give a significant elimination of the investigated compounds due to hydrolysis and much lower siloxanes concentrations in the biogas of the following anaerobic digester. Similar pre-digester reactors operated at short retention times and neutral pH have been used in the

literature to improve sludge solubilisation and overall biogas production of the system. Having in mind the high sorption coefficients of longer siloxanes (Table 3), sorption could also be an important mechanism governing the fate of these compounds in this pre-digester tank.

To test this hypothesis, a two-stage system was considered consisting of a pre-digester (Tank 1, HRT: 3 d) and an anaerobic digester (Tank 2), operating at the same conditions described before (Table S7), and the fate of investigated siloxanes was predicted for mesophilic and thermophilic conditions. Equation 6 was used for both reactors with the difference that volatilization was considered negligible at the pre-digester due to its low HRT which prevent methanogens from developing. The results presented in Figure 3 reveal that more than 92% of D3 and an important part of D4 is expected to be removed via hydrolysis in the pre-digester tank under both mesophilic and thermophilic conditions. Additionally, the major part of D5 and D6 (>98%) is seen to accumulate in sludge in Tank 1, while significant sorbed amounts are also predicted for D4 under mesophilic (69.5%) and thermophilic conditions (47.5%). Due to the aforementioned mechanisms occurring in the pre-digester, the amount of siloxanes that volatilize in the anaerobic digester is much smaller. As a result, just 4.0% of D3 and 13.4% of D4 can be predicted to be transferred to the biogas in the mesophilic reactor, while even lower fractions are predicted for thermophilic reactor. Much lower fractions are also predicted for D5 and D6 in biogas (Figure 3). Back-calculation of these results shows that the expected concentrations of D3, D4 and D5 in the biogas are one or two order of magnitude lower than those estimated in single-stage system, ranging between $4.0 \mu\text{g m}^{-3}$ (D5, mesophilic conditions) and $97.3 \mu\text{g m}^{-3}$ (D4, mesophilic conditions).

4. Conclusions

D3 was rapidly hydrolysed at all tested temperatures, while hydrolysis of other siloxanes was slower. The increase of temperature accelerated hydrolysis of investigated siloxanes. Sorption was different among the investigated compounds with logKd values ranging from 1.46 (D3) to 3.84 (D5). The fate of cyclic siloxanes during single-stage anaerobic digestion is affected mainly by volatilization and transfer to biogas. Mass balances predict that the use of a pre-digester in a two-stage system could increase hydrolysis of D3, D4 and sorption of D4, D5 and D6 resulting in mitigation of the significant problem of siloxanes in biogas produced from sewage sludge.

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6. References

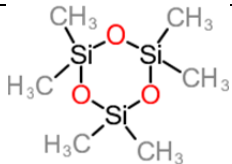
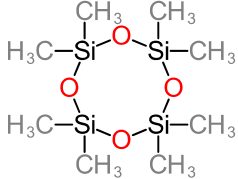
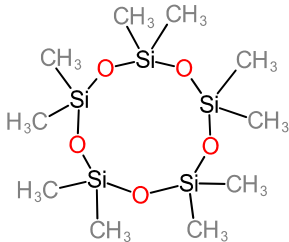
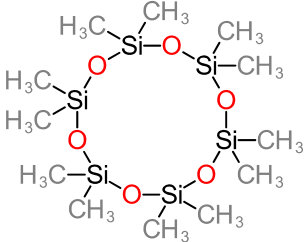
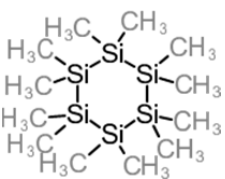
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Table 1. Abbreviations, structures, molecular weights and physico-chemical properties of target cyclic siloxanes.

Compound	Structure	Molecular Weight (g mol ⁻¹)	^a Vapour pressure at 25 °C (Pa)	^a Water solubility at 25 °C (mg/L)	^a Log K _{OW}	^b Log K _{AW}	^c Log K _{OC}
D3		222	1146.6	1.56	3.85	-	3.55
D4		297	131.99	0.056	4.45	2.69 ^b	4.17
D5		371	23.198	0.017	5.2	3.13 ^b	4.60
D6		445	3.999	0.005	5.86	3.01 ^b	5.08
D6_silane		349	-	-	-	-	-

^aPalczewska-Tulińska and Oracz, 2005; ^bXu and Kropscott, 2012; ^cAllen et al., 1997

Table 2. First order kinetics ($k_{\text{hydrolysis}}$), half-life values and R^2 of target compounds, calculated during hydrolysis experiments under different experimental temperature conditions. The 95% confidence intervals of measured values are given in parenthesis.

Compound	Temperature (°C)	$k_{\text{hydrolysis}}$ (d ⁻¹)	Half-life (d)	R^2
D3	4 °C	2.5 (± 0.90)	0.27 (±0.07)	0.85
	20 °C	4.1 (± 1.67)	0.17 (± 0.05)	0.83
	35 °C	7.2 (± 2.74)	0.10 (± 0.03)	0.88
	55 °C	10.3 (± 2.80)	0.07 (± 0.01)	0.96
D4	4 °C	0.05 (± 0.02)	15.2 (± 4.3)	0.63
	20 °C	0.17 (± 0.05)	4.1 (± 0.9)	0.88
	35 °C	0.43 (± 0.08)	1.6 (± 0.3)	0.97
	55 °C	1.58 (± 0.20)	0.4 (± 0.05)	0.99
D5	4 °C	0.03 (± 0.01)	21.8 (± 5.0)	0.71
	20 °C	0.08 (± 0.02)	8.2 (± 1.6)	0.87
	35 °C	0.10 (± 0.03)	6.7 (± 1.4)	0.87
	55 °C	0.11 (± 0.03)	6.2 (± 1.4)	0.85
D6	4 °C	0.03 (± 0.02)	24.3 (± 9.4)	0.42
	20 °C	0.06 (± 0.02)	11.4 (± 3.0)	0.73
	35 °C	0.08 (± 0.02)	9.2 (± 2.2)	0.82
	55 °C	0.05 (± 0.02)	14.8 (± 3.9)	0.78
D6_silane	4 °C	0.01 (±0.01)	48.4 (± 17.1)	0.42
	20 °C	0.06 (± 0.02)	12.5 (± 3.1)	0.73
	35 °C	0.10 (± 0.02)	7.1 (± 1.3)	0.91
	55 °C	0.03 (± 0.02)	20.0 (± 6.1)	0.58

Table 3. Average log K_d values for siloxanes in anaerobically digested sludge (35 °C) and comparison with literature data. Standard deviations of measured log K_d values are given in parenthesis.

Target Analytes	Log K_d (L·Kg ⁻¹)					
	Anaerobically Digested Sludge ^a	Primary Sludge ^b	Secondary Sludge ^c	Primary Sludge ^c	Secondary Sludge ^c	Aerobic Activated Sludge ^d
D3	1.46 ± 0.95	1.83	2.15	-	-	2.23
D4	2.32 ± 1.85	3.05	3.40	3.30	3.64	1.94
D5	3.84 ± 3.42	3.64	4.42	3.41	5.50	2.46
D6	3.41 ± 2.75	3.49	3.95	3.53	5.57	2.17
D6_silane	2.82 ± 2.34					

^acurrent study; ^bBletsou et al., 2013; ^cWang et al., 2015; ^dXu et al., 2013

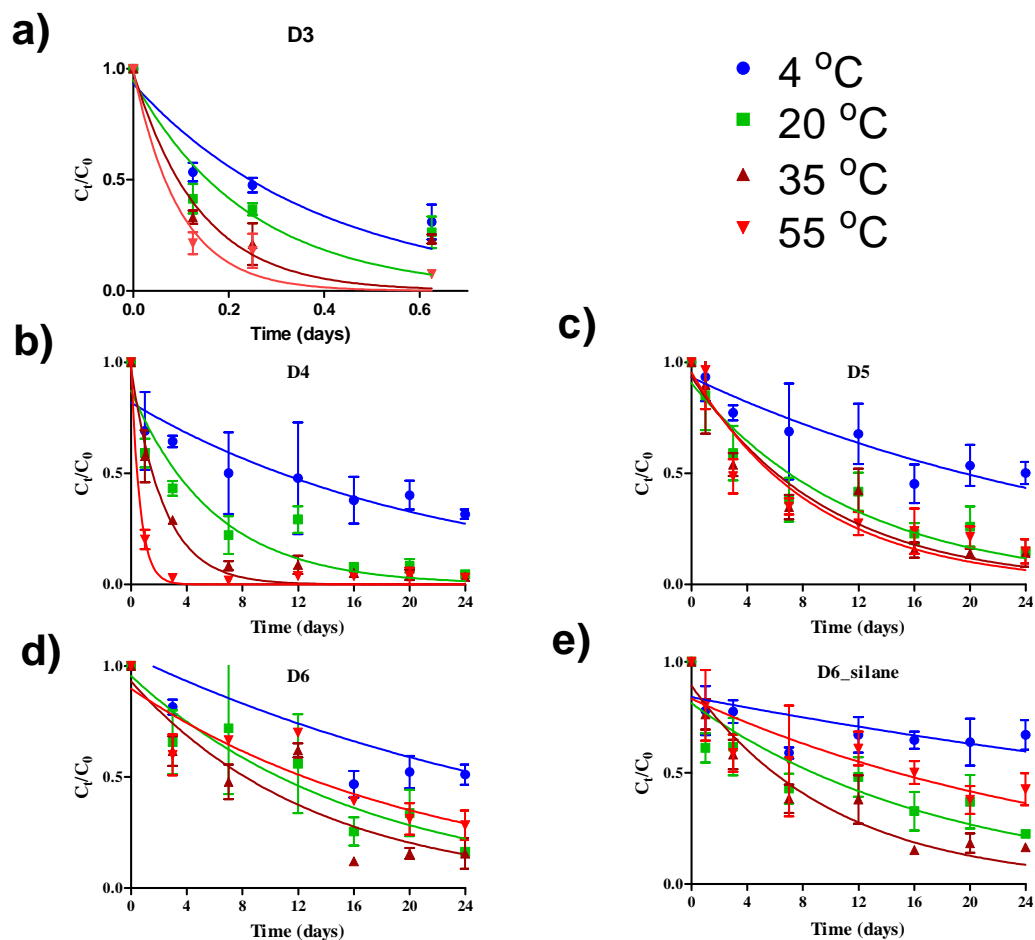


Fig. 1. Reduction of target siloxanes during hydrolysis experiments conducted at different temperatures; a) D3 (total duration of the experiment: 15 hours), b-e) D4, D5, D6, D6_silane (total duration of the experiment: 24 days).

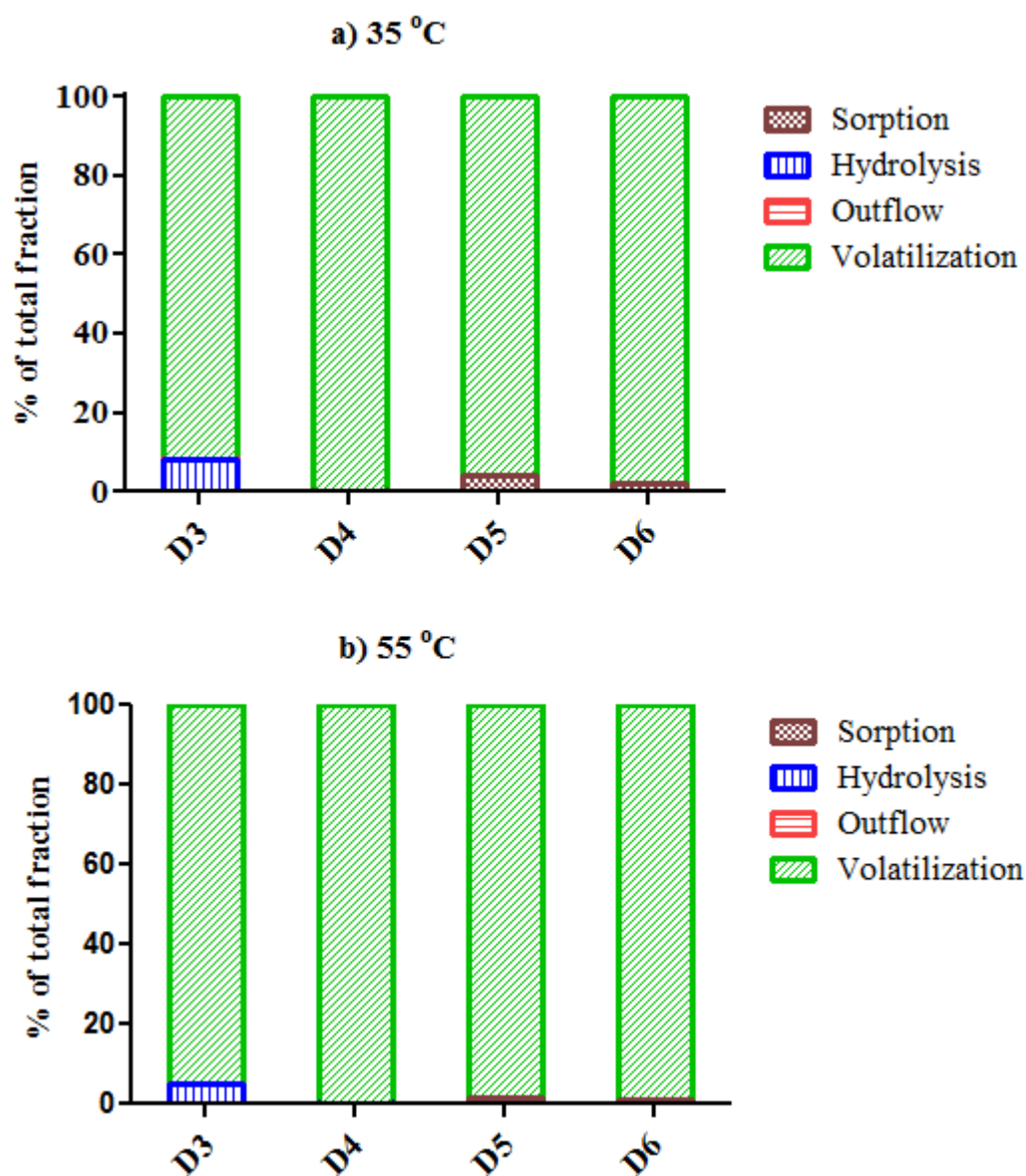
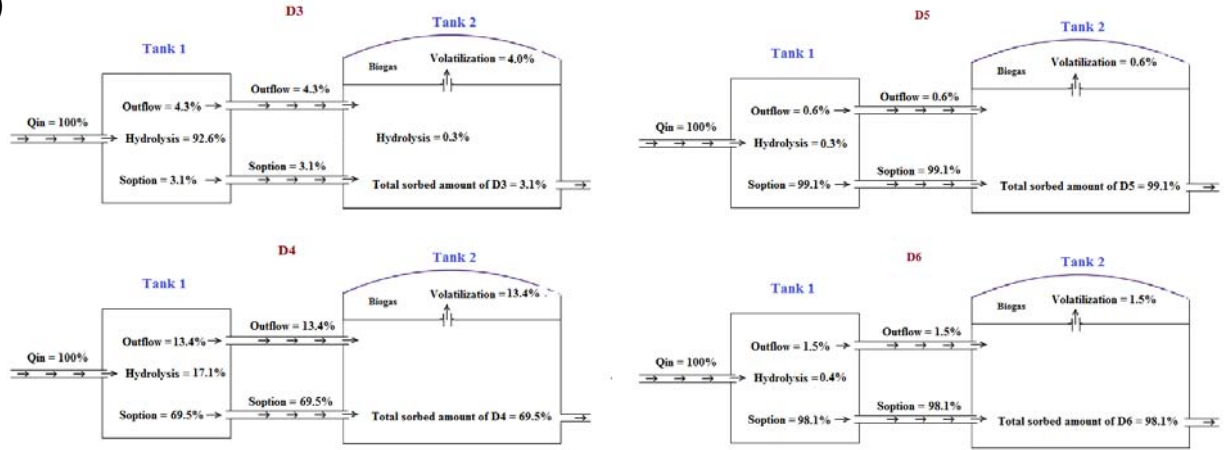


Fig. 2. Fate of target siloxanes during one-stage (a) mesophilic and (b) thermophilic anaerobic sludge digestion.

a)



b)

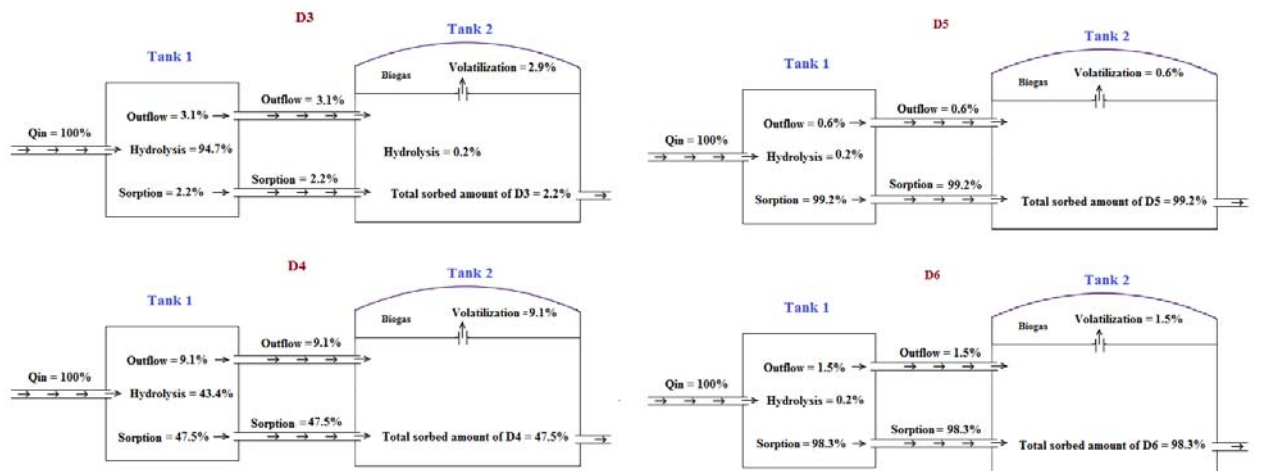


Fig. 3. Fate of D3, D4, D5 and D6 during (a) mesophilic and (b) thermophilic two-stage anaerobic sludge digestion (Tank 1: pre-digester; Tank 2: anaerobic digester).